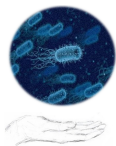


Dynamics of antibiotic stress response by a Type II toxin-antitoxin system

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Outline

- 1 Type II introduction
 - System regulation
 - Predictions of spontaneous persister formation
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- 3 Conclusion and discussion
 - Modelling
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Regulation of Type II TA systems

- Toxin ("T") and antitoxin ("A") are expressed together.
- T generally inhibits translation and leads to cell growth arrest.
- T forms complex with A which prevents T toxicity.
- Complex formation is cooperative.
- Complex of A with T binds to their promoter.
- There is non-linear negative feedback.
- Details may vary for different TA systems.

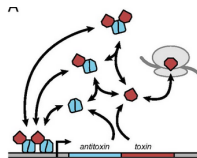


Figure: PMID 23781105: Regulation and complex formation in Type II TA

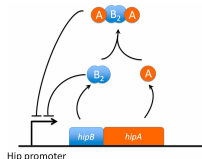


Figure: PMID 24344277: A variation



Positive feedback due to toxin inhibition of cell growth

- T inhibits cell growth. Slower cell growth leads to lower protein dilution. This leads to smaller effective T degradation.
- It is assumed that several TA systems in the genome may act cooperatively! Therefore, in addition to positive, there may also be a nonlinear positive feedback in the system.

Bistability may emerge!

High toxin state may correspond to persisters.

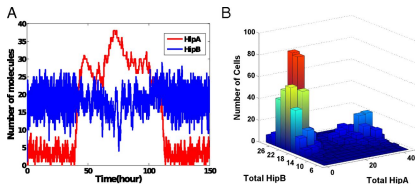


Figure: PMID 24344277: Modelling predicts spontaneous persister formation



kacAT configuration and experimental observations

kacAT properties

- kacAT architecture generally similar to other Type II systems
- Large binding cooperativity where complex is formed by 2T and 4A
- Strong system response to stress
- However, no induction of spontaneous persistence!
- Antitoxin is degraded in response to stress

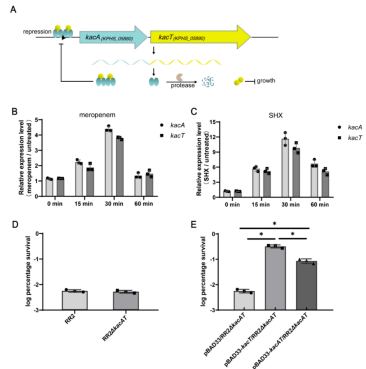


Figure: *kacAT* regulation and experimental results [1]

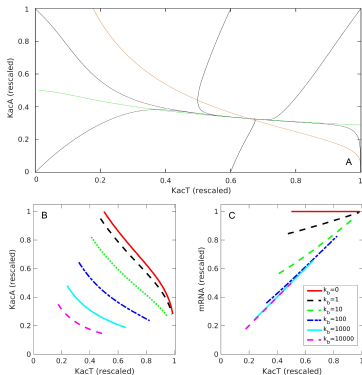
Mathematical model of the system dynamics [1]:

$$\frac{dA}{d\tau} = \frac{2k\phi}{1 + K_b A^4 B^2} - (1 + \delta\lambda)A - 4A^4 B^2$$

$$\frac{dB}{d\tau} = \frac{k\phi}{1 + K_b A^4 B^2} - B - 2A^4 B^2$$

Model explains:

- Reduction of the [KacA]:[KacT] ratio upon antibiotic application.
- Large increase in *kacAT* transcripts induced by antibiotics.
- *KacAT* overexpression induces antibiotic stress tolerance, whereas *kacAT* deletion does not affect this tolerance.





Persister emergence model assumption

- Why *kacAT* deletion does not lead to spontaneous persister formation despite previous theoretical predictions?
- Theoretical studies assume cooperative (joint) action of several TAs.

Only one system in the same family as *kacAT* strongly preferred!

Also, statistically significant negative correlations between different clades for which experiments indicate their cross-talk [2]

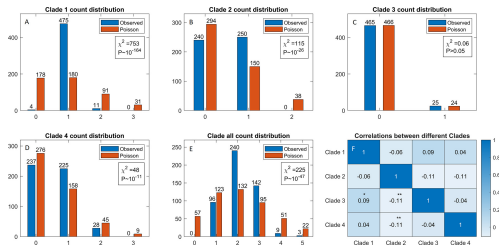


Figure: Statistical analysis of GNAT-RHH TA pairs in *K. pneumoniae* strains.

Conclusions

Developed mechanistic model of kacAT system dynamics

- kacAT regulation leads to monostable dynamics.
- Can explain all experimental results.
- Explains that kacAT delation does not affect cell growth.

Predicted and analyzed all GNAT-RHH TA pairs in *K. pneumoniae*

- Multiple loci instances (or absence of loci) are strongly disfavoured.
- Crosstalk between loci also appears disfavoured.
- This may contribute to the observed absence of spontaneous persister formation in kacAT.

Outlook: Can kacAT lead to antibiotic induced persister formation?

References

- [1] Peifei Li, Ying-Xian Goh, Bojana Ilic, Cui Tai, Zixin Deng, Zhaoyan Chen, Marko Djordjevic, and Hong-Yu Ou. Antibiotic-induced degradation of antitoxin enhances the transcription of acetyltransferase-type toxin-antitoxin operon. *The Journal of Antimicrobial Chemotherapy*, 78(4).
- [2] Ying-Xian Goh, Peifei Li, Meng Wang, Marko Djordjevic, Cui Tai, Hui Wang, Zixin Deng, Zhaoyan Chen, and Hong-Yu Ou. Comparative Analysis of Diverse Acetyltransferase-Type Toxin-Antitoxin Loci in *Klebsiella pneumoniae*. *Microbiology Spectrum*, 10(4).

Acknowledgments

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